

# Comparative Evaluation of Molecular vs. Conventional Techniques in the Diagnosis of Bloodstream Infections

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## Abstract

**Background:** Bloodstream infections (BSIs) are a major cause of morbidity and mortality, requiring prompt and accurate diagnosis for effective management. This study compared the performance of molecular diagnostic methods with conventional blood culture techniques in detecting BSIs in a tertiary hospital setting.

**Methods:** A prospective observational study was conducted over 12 months. Blood samples from 500 patients with suspected BSIs were analyzed using conventional blood culture and molecular diagnostics (FilmArray Blood Culture Identification Panel). Key parameters, including sensitivity, specificity, time to detection, and therapy modification rates, were evaluated.

**Results:** Molecular diagnostics demonstrated higher sensitivity (95% vs. 85%) and comparable specificity (92% vs. 90%) compared to conventional methods. The median time to detection was significantly reduced (2–6 hours vs. 24–72 hours), leading to higher therapy modification rates (85% vs. 60%). Molecular diagnostics also identified pathogens missed by conventional methods, particularly fastidious organisms like *Candida* spp.\* and *Pseudomonas aeruginosa*.\*

**Conclusion:** Molecular diagnostic methods significantly improved the speed and accuracy of BSI detection, influencing timely therapy modifications and enhancing patient outcomes. However, high costs and lack of antimicrobial susceptibility profiling remain challenges to widespread adoption.

**Keywords:** Bloodstream Infections, Molecular Diagnostics, Blood Culture, Sensitivity, Specificity, Pathogen Detection, Antimicrobial Stewardship.

## Introduction

Bloodstream infections (BSIs) represent a critical healthcare challenge, contributing significantly to morbidity and mortality, particularly in critically ill patients. Early and accurate diagnosis is paramount for initiating effective antimicrobial therapy and improving patient outcomes (Singer et al., 2016). Traditionally, the detection of BSIs has relied on blood culture techniques, which, although considered the gold standard, are hindered by prolonged turnaround times (24–72 hours or longer) and limited sensitivity, especially for certain pathogens like fastidious or intracellular organisms (Kirn & Weinstein, 2013).

In recent years, molecular diagnostic technologies have emerged as promising tools to address the limitations of conventional blood culture methods. Techniques such as polymerase chain reaction (PCR), multiplex PCR panels, and next-generation sequencing (NGS) enable direct identification of microbial DNA or RNA from blood samples, providing faster and more sensitive results (Ecker et al., 2010). For example, studies have demonstrated that molecular diagnostics can reduce the time to pathogen identification by several hours to days, which is critical for initiating targeted antimicrobial therapy and improving patient survival rates (Thaden et al., 2022; Babafemi et al., 2017).

Despite these advancements, the implementation of molecular methods into routine clinical practice faces challenges, including high costs, the need for specialized equipment, and the inability of some molecular tests to determine antimicrobial susceptibility profiles, which are crucial for guiding therapy (Cohen et al., 2015). Additionally, the potential for detecting non-viable pathogens or contaminants further complicates the interpretation of results (Dark et al., 2012).

This study aims to conduct a comparative evaluation of molecular and conventional diagnostic techniques for BSIs, focusing on parameters such as time to detection, sensitivity, specificity, and clinical impact. By elucidating the strengths and limitations of each approach, this research seeks to provide evidence-based recommendations for optimizing the diagnostic management of BSIs.

### **Literature Review**

Bloodstream infections (BSIs) are a major cause of morbidity and mortality worldwide, with effective diagnosis being essential for guiding targeted antimicrobial therapy. Conventional blood culture methods have been the cornerstone of BSI diagnosis for decades, offering the advantage of detecting viable organisms and enabling antimicrobial susceptibility testing. However, these methods are hampered by prolonged turnaround times (24–72 hours) and reduced sensitivity, especially in patients already receiving antibiotics (Kirn & Weinstein, 2013).

Molecular diagnostic techniques, such as polymerase chain reaction (PCR), multiplex PCR panels, and next-generation sequencing (NGS), have emerged as promising tools for improving the detection of pathogens in BSIs. These methods allow for rapid and direct identification of microbial DNA or RNA from blood samples, significantly reducing the time to pathogen identification (Thaden et al., 2022). For example, multiplex PCR panels can identify a range of pathogens within hours, with reported sensitivities exceeding 80% and specificities over 90% for common bloodstream pathogens (Babafemi et al., 2017).

Recent studies have highlighted the clinical benefits of molecular diagnostics in managing BSIs. The use of molecular techniques has been associated with earlier initiation of targeted therapy, reduced hospital stays, and decreased mortality rates. In a systematic review, rapid molecular diagnostics were shown to reduce the time to appropriate antimicrobial therapy by up to 24 hours, contributing to improved patient outcomes (Thaden et al., 2022). Furthermore, molecular methods can detect pathogens that are difficult to culture, such as fastidious or slow-growing organisms (Ecker et al., 2010).

Despite these advantages, several challenges limit the widespread adoption of molecular diagnostics in routine clinical practice. High costs, the need for specialized equipment, and the requirement for trained personnel present significant barriers, particularly in resource-limited settings (Cohen et al., 2015). Additionally, while molecular methods provide rapid identification, they often lack the ability to deliver

detailed antimicrobial susceptibility profiles, which remain crucial for tailoring treatment regimens (Dark et al., 2012).

Overall, while molecular diagnostics offer substantial advantages in the detection and management of BSIs, their integration into clinical workflows must address these practical and financial challenges. This review seeks to critically assess the comparative performance of molecular and conventional diagnostic methods, focusing on sensitivity, specificity, time to detection, and clinical outcomes.

## Methodology

This study was conducted at a tertiary care hospital over a 12-month period to compare the diagnostic performance of molecular and conventional techniques in identifying bloodstream infections (BSIs). The hospital's microbiology laboratory served as the central facility for processing and analyzing samples.

### Study Design

This was a prospective, observational study designed to evaluate the sensitivity, specificity, time to pathogen detection, and clinical impact of molecular diagnostic techniques compared to traditional blood culture methods.

### Study Population

Patients admitted to the hospital with suspected bloodstream infections were enrolled in the study. Inclusion criteria were:

1. Patients aged  $\geq 18$  years.
2. Clinical suspicion of BSI, based on signs such as fever, chills, or hypotension.
3. Collection of blood samples for diagnostic purposes.

Exclusion criteria included:

1. Patients who had already initiated broad-spectrum antibiotic therapy without prior blood sampling.
2. Inadequate or improperly collected blood samples.

### Sample Collection

For each patient, two sets of blood samples were collected:

1. Blood Culture: Each set consisted of aerobic and anaerobic bottles, with a volume of 10 mL per bottle. Samples were processed using the BacT/ALERT system (bioMérieux).
2. Molecular Diagnostics: An additional 5 mL of blood was collected in EDTA tubes for molecular testing.

### Laboratory Procedures

#### 1. Conventional Blood Culture

Blood culture bottles were incubated in the automated BacT/ALERT system. Positive cultures were subjected to Gram staining, followed by identification using biochemical tests or matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS). Antimicrobial susceptibility testing was performed using the VITEK 2 system (bioMérieux).

#### 2. Molecular Diagnostics

Molecular analysis was performed using the FilmArray Blood Culture Identification Panel (BioFire

Diagnostics). The panel identifies over 20 common bloodstream pathogens and resistance markers (e.g., *mecA*, *vanA/B*). Results were generated within 2 hours from DNA extraction to reporting.

#### Data Collection

Data were collected on the following variables:

- Time to detection (from sample collection to reporting).
- Pathogen identification and resistance profiling results.
- Concordance between molecular and conventional methods.
- Changes in clinical management (e.g., initiation or modification of antimicrobial therapy) based on diagnostic results.
- Patient outcomes, including length of hospital stay and 30-day mortality.

#### Statistical Analysis

The diagnostic performance of each method was evaluated based on:

- Sensitivity and Specificity: Calculated by comparing results to a composite reference standard (conventional culture, molecular detection, and clinical findings).
- Time to Detection: Median time was compared using the Wilcoxon rank-sum test.
- Clinical Impact: Differences in therapy modification rates and outcomes were assessed using chi-square tests or logistic regression.

#### Ethical Considerations

The study was approved by the ethics committee. Written informed consent was obtained from all participants before sample collection. Patient data were anonymized to ensure confidentiality.

#### Findings

This study assessed the diagnostic performance of molecular and conventional methods in detecting bloodstream infections (BSIs). The findings are presented below.

#### Key Diagnostic Parameters

The comparison of sensitivity, specificity, median time to detection, and therapy modification rates is summarized in Table 1.

**Table 1: Key Findings of Diagnostic Methods for BSIs**

| Parameter                 | Conventional Methods | Molecular Methods |
|---------------------------|----------------------|-------------------|
| Sensitivity               | 85%                  | 95%               |
| Specificity               | 90%                  | 92%               |
| Median Time to Detection  | 24–72 hours          | 2–6 hours         |
| Therapy Modification Rate | 60%                  | 85%               |

#### Pathogen Detection

The detection rates of common pathogens using the two methods are shown in Table 2. Molecular methods demonstrated higher detection rates, particularly for fastidious and hard-to-culture organisms.

**Table 2: Pathogen Detection Rates: Conventional vs Molecular Methods**

| Pathogen                      | Detected by Conventional Methods (%) | Detected by Molecular Methods (%) |
|-------------------------------|--------------------------------------|-----------------------------------|
| <i>Escherichia coli</i>       | 80                                   | 95                                |
| <i>Staphylococcus aureus</i>  | 78                                   | 90                                |
| <i>Klebsiella pneumoniae</i>  | 85                                   | 92                                |
| <i>Pseudomonas aeruginosa</i> | 70                                   | 88                                |
| <i>Candida spp.</i>           | 60                                   | 85                                |

### Key Observations

1. Time to Detection: Molecular methods significantly reduced the time to pathogen identification, with results available within 2–6 hours compared to 24–72 hours for conventional methods.
2. Sensitivity and Specificity: Molecular diagnostics demonstrated higher sensitivity (95%) and comparable specificity (92%) compared to conventional blood cultures.
3. Pathogen Identification: Molecular methods detected pathogens in cases where conventional methods yielded negative results, particularly for *Candida spp.* and *Pseudomonas aeruginosa*.
4. Clinical Impact: Therapy modification based on molecular results was more frequent (85%) compared to conventional methods (60%), leading to improved antimicrobial targeting.

### Discussion

This study compared the performance of molecular diagnostic techniques and conventional blood culture methods in the detection of bloodstream infections (BSIs). The findings reveal significant advantages of molecular diagnostics, particularly in terms of sensitivity, specificity, and time to pathogen detection, which have critical implications for clinical decision-making and patient outcomes.

#### Sensitivity and Specificity

Molecular diagnostic methods demonstrated higher sensitivity (95%) and comparable specificity (92%) compared to conventional blood culture techniques (sensitivity 85%, specificity 90%). These results align with previous studies, which have shown that molecular methods are particularly effective in detecting pathogens in cases where blood cultures may fail, such as in patients receiving prior antibiotic therapy or infections caused by fastidious organisms (Thaden et al., 2022; Babafemi et al., 2017). For example, molecular diagnostics detected higher rates of *Candida spp.* and *Pseudomonas aeruginosa*, both of which are challenging to identify using conventional methods. This improved sensitivity can lead to earlier and more accurate diagnoses, reducing the risk of delayed or inappropriate therapy.

#### Time to Detection

One of the most significant advantages of molecular diagnostics was the reduced time to detection. Molecular methods provided results within 2–6 hours, whereas conventional blood cultures required 24–72 hours. This reduction in diagnostic time is critical for BSIs, where every hour of delayed treatment increases the risk of mortality (Kumar et al., 2006). Rapid identification allows for earlier initiation of targeted antimicrobial therapy, which not only improves patient outcomes but also reduces the length of hospital stay and healthcare costs.

### Therapy Modification

The study found that therapy modification rates were significantly higher for molecular diagnostics (85%) compared to conventional methods (60%). This indicates that molecular methods not only expedite diagnosis but also influence clinical decision-making. By providing rapid identification of pathogens and key resistance markers (e.g., *mecA*, *vanA/B*), molecular diagnostics enable clinicians to tailor antimicrobial therapy more effectively. This finding is consistent with previous research emphasizing the role of molecular tools in improving antimicrobial stewardship programs (Cohen et al., 2015).

### Limitations of Molecular Diagnostics

Despite their advantages, molecular diagnostic methods have limitations that may impact their integration into routine clinical practice. These include higher costs, the need for specialized equipment, and the potential for detecting non-viable pathogens or contaminants, which may lead to overdiagnosis or unnecessary treatment (Dark et al., 2012). Additionally, molecular diagnostics often lack comprehensive antimicrobial susceptibility testing, requiring supplementary culture-based methods for resistance profiling.

### Clinical Implications

The findings of this study underscore the potential of molecular diagnostics to transform the management of BSIs in tertiary hospital settings. By reducing diagnostic time, improving pathogen detection, and enabling timely therapy modifications, these methods can significantly enhance patient outcomes. However, their implementation must be balanced against cost considerations and the need for complementary culture-based methods.

### Future Directions

Future research should focus on cost-effectiveness analyses of molecular diagnostics in real-world clinical settings and the development of integrated platforms that combine rapid pathogen detection with antimicrobial susceptibility testing. Additionally, efforts should be made to address barriers to implementation, particularly in resource-limited settings.

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